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101.211-24u – without *Taq* **polymerase** General "Instructions for Use"

IFU-02 Rev. No. 01 can be downloaded from

Lot No.: 83G Lot-specific information www.olerup-ssp.com

Olerup SSP® DQB1*05

Product number: 101.211-24u – without *Taq* polymerase

Lot number: 83G

Expiry date: 2012-February-01

Number of tests: 24 Number of wells per test: 6

Storage - pre-aliquoted primers: dark at -20°C

- PCR Master Mix: -20°C
- Adhesive PCR seals RT
- Product Insert RT

This Product Description is only valid for Lot No. 83G

CHANGES COMPARED TO THE PREVIOUS OLERUP SSP® DQB1*05 LOT

The DQB1*05 primer set, specificity and interpretation tables are unchanged compared to the previous *Olerup* SSP® DQB1*05 lot (Lot No. 42F).

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PRODUCT DESCRIPTION

DQB1*05 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1*0501 to DQB1*0505 alleles.

Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.

PLATE LAYOUT

Each test consists of 6 PCR reactions in an 8 well PCR plate. Wells 7 and 8 are empty.

1 2 3 4 5 6 empty empty

The 8 well cut PCR plate is marked with 'DQ5' in silver gray ink.

Well No. 1 is marked with the Lot No. '83G'.

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded.

The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each 8 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Only the DQB1*05 alleles will be amplified by the DQB1*05 subtyping kit. Thus, the interpretation of DQB1*05 subtypings is not influenced by other groups of DQB1 alleles or the DQB2 and DQB3 genes.

UNIQUELY IDENTIFIED ALLELES

All the DQB1*05 alleles, i.e. **DQB1*0501 to DQB1*0505**, recognized by the HLA Nomenclature Committee in January 2010¹ will give rise to unique amplification patterns by the primers in the DQB1*05 subtyping kit.

The DQB1*05 subtyping kit cannot distinguish the DQB1*050101 and DQB1*050102 alleles or the DQB1*050301 and DQB1*050302 alleles.

¹DQB1 alleles listed on the IMGT/HLA web page 2010-January-15, release 2.28.0, www.ebi.ac.uk/imgt/hla.

CE

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RESOLUTION IN HOMO- AND HETEROZYGOTES

The 5 DQB1*05 alleles generate 6 different amplification patterns, as the DQB1*050201 and DQB1*050202 alleles generate different amplification patterns. These can be combined in 21 homozygous and heterozygous combinations. Eight of these genotypes do not give rise to unique amplification patterns.

```
050202,0505 = 0503,0505
          050201,050202 = 050201,0503 = 050202,050202 =
          050202,0503
         050201,0505 = 0505,0505
+-+-+
```

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101.211-24u - without *Taq* polymerase

Lot No.: 83G

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SPECIFICITY TABLE

DQB1*05 SSP subtyping

Specificities and sizes of the PCR products of the 6 primer mixes used for DQB1*05 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified DQB1*05 ³ alleles
1	225 bp	515 bp	*050101-0505
2	135 bp	430 bp	*050101, 050102
3 ⁴	120 bp	430 bp	*050201, 050202, 0505
4 ⁴	95 bp	515 bp	*050202-050302
5 ⁴	120 bp	430 bp	*0504
6	185 bp	430 bp	*0505

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQB1*05 SSP typings.

When the primers in a primer mix can give rise to specific PCR products of more than one length this is indicated if the size difference is 20 base pairs or more. Size differences shorter than 20 base pairs are not given. For high resolution SSP kits the respective length of the specific PCR product(s) of the alleles amplified by these primer mixes are given.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherit feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1*05 subtyping.

In addition, well number 4 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to allow kit identification.

In the presence of a specific amplification the intensity of the control band often decreases.

³For several DQB alleles only partial second exon nucleotide sequences are available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. We assume that unknown sequences of codons 87 to 92 are conserved within allelic groups.

⁴Specific PCR fragments shorter than 125 base pairs have a lower intensity than longer PCR bands.

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INTERPRETATION TABLE							
	DQB1*	05 SSF	subty	ping			
Amplific	cation pa	atterns o	f the DQ	B1*05 al	leles		
	Well						
	1	2	3	4	5	6	
Length of spec.	225	135	120	95	120	185	
PCR product							
Length of int.	515	430	430	515	430	430	
pos. control ¹	pos. control ¹						
5'-primer ²	26(173)	26(173)	30(184)	30(184)	30(184)	38(209)	
	^{5'} -ggg ^{3'}	^{5'} -ggg ^{3'}	^{5'} -gAC ^{3'}	^{5'} -gAC ^{3'}	^{5'} -gAT ^{3'}	^{5'} -CgC ^{3'}	
3'-primer ³	87(356)	57(266)	57(265)	47(237)	57(265)	86(353)	
	^{5'} -ggT ^{3'}	^{5'} -CAA ^{3'}	^{5'} -gCT ^{3'}	^{5'} -CgA ^{3'}	^{5'} -gCT ^{3'}	^{5'} -ACg ^{3'}	
Well No.	1	2	3	4	5	6	
DQB1 allele							
*050101, 050102	1	2					
*050201	1		3				
*050202	1		3	4			
*050301, 050302	1			4			
*0504	1				5		
*0505	1		3			6	
DQB1 allele							
Well No.	1	2	3	4	5	6	

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The two different control primer pairs give rise to either an internal positive control band of 430 base pairs, for most wells, or a band of 515 base pairs, for some wells.

Well number 1 contains the primer pair giving rise to the longer, 515 bp, internal positive control band in order to help in the correct orientation of the DQB1*05 subtyping.

In addition, well number 4 contains the primer pair giving rise to the longer, 515 bp, internal

positive control band in order to allow kit identification. ²The codon, and in parenthesis the nucleotide, in the 2nd exon, matching the specificitydetermining 3'-end of the primer is given. Codon and nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is

given. ³The codon, and in parenthesis the nucleotide, in the 2nd exon, matching the specificitydetermining 3'-end of the primer is given in the anti-sense direction. Codon and nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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CELL LINE VALIDATION SHEET										
DQB1*05 SSP subtyping kit										
		Well								
					1	2	3	4	5	6
					_					
				Production No.	200969601	200969602	200969603	200969604	200969605	200969606
				L C	696	696	696	696	696	969
				<u>0</u> 0	800	8	8	8	8	00
	ILIVA	/C cell line	D/		2	7	7	7	2	2
1	9001		*0501	QB1	+	+	-	-	-	-
2		LK707	*0601	*0202	-	-	-	-	-	_
3		E4181324	*0601	0202	-	-	-	-	-	-
4		GU373	*0201		-	-	-	-	-	-
5		KAS011	*0502		+	-	+	-	-	-
6	9353		*0302	*0601	-	-	 	-	-	-
7	9020		*0201		-	-	-	-	-	-
8	9025		*0301		-	-	-	-	-	-
9	9026	YAR	*0302		·	-	-	-	-	-
10	9107	LKT3	*0401		٠	-	-	-	-	-
11	9051	PITOUT	*0202		-	-	-	-	-	-
12	9052		*0303		•	-	-	-	-	-
13		JESTHOM	*0501	1	+	+	-	-	-	-
14		OLGA	*0402	1	-	-	-	-	-	-
15	9075		*0303	1	_	-	-	<u> </u>	<u> </u>	-
16		SWEIG007	*0301		-	-	-	<u>-</u>	<u>-</u>	-
17		CTM3953540	*0201	*0603	•	-	-	-	-	-
18		32367	*0602	*0202	•	-	-	-	-	-
19		BM16	*0301		-	-	-	-	-	-
20 21		SLE005 AMALA	*0604	+	-	-	-	-	-	-
22		KOSE	*0301 *0503	*0604	÷	Η-	-	+	H	
23	9124		*0503	*0601	+	-	-	+	-	-
24		JBUSH	*0301	0001	Ė.	-	-	Ė	-	-
25		IBW9	*0202		-	-	-	-	-	-
26		WT49	*0201		-	-	-	-	-	-
27		CH1007	*0401	*0501	+	+	-	-	-	-
28	9320	BEL5GB	*0202	*0301	-	-	-	-	-	-
29	9050	MOU	*0202		_				-	-
30	9021	RSH	*0402		•	-	-	-	-	-
31		DUCAF	*0201		٠	-	-	-	-	-
32	9297		*0301	1	•	-	-	-	-	-
33		MT14B	*0302	1	-	-	-	-	-	-
34		DHIF	*0301		-	-	-	-	-	-
35		SSTO	*0305		-	-	-	-	-	-
36 37		KT17	*0302	1	-	<u>-</u>	<u>-</u>	-	-	-
38	9065	HHKB	*0603		H	Ė	Ė	Ė	Ė	
39	9099		*0301 *0201	*0301	Ë	-	-	-	-	-
40		WHONP199	*0201	*0303	-	-	-	-	-	
41		H0301	*0609	0000	-	-	-	-	-	
42		TAB089	*0601		-	-	-	-	-	-
43		T7526	*0303	1	-	-	-	-	-	-
44	9057		*0503		+	-	-	+	-	-
45		SHJO	*0202	1	-	-	-	-	-	-
46		SCHU	*0602		-	-	-	-	-	-
47		TUBO	*0301		-	-	-	-	-	-
48		TER-ND	*0501		+	+	-	-	-	

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CERTIFICATE OF ANALYSIS

Olerup SSP® DQB1*05 SSP

Product number: 101.211-24u – without *Taq* polymerase

Lot number: 83G

Expiry date: 2012-February-01

Number of tests: 24 Number of wells per test: 6

Well specifications:

Well No.	Production No.
1	2009-696-01
2	2009-696-02
3	2009-696-03
4	2009-696-04
5	2009-696-05
6	2009-696-06

The specificity of each primer solution of the kit has been tested against 48 well characterized IHWC cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 5 and 6 were available. The specificities of the primers in theise primer solutions were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer.

Results: No false positive or false negative amplifications were obtained.

Date of approval: 2010-February-16

Approved by:

Quality Control, Supervisor

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101.211-24u – without *Taq* polymerase

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Declaration of Conformity

Product name: Olerup SSP® DQB1*05

Product number: 101.211-24u

Lot number: 83G

Intended use: DQB1*05 resolution histocompatibility testing

Manufacturer: Olerup SSP AB

Hasselstigen 1

SE-133 33 Saltsjöbaden, Sweden

Phone: +46-8-717 88 27 **Fax:** +46-8-717 88 18

We, *Olerup* SSP AB, hereby declare that this product, to which this Declaration of Conformity relates is in conformity with the following Standard(s) and other normative document(s) ISO 9001:2008 and ISO 13485:2003, following the provisions of the 98/79/EC Directive on *in vitro* diagnostic medical devices, Annex II List B, conformity assessed using Annex IV, as transposed into the national laws of the Member States of the European Union.

The Technical Documentation File is maintained at *Olerup* SSP AB, Hasselstigen 1, SE-133 33 Saltsjöbaden, Sweden.

The Authorized Representative located within the Community is: *Olerup* SSP AB.

Saltsjöbaden, Sweden 2010-February-16

Olle Olerup Managing Director DQB1*05 Product Insert Page 9 of 12 101.211-24u – without *Taq* polymerase General "Instructions for Use"

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